

Chronic Lymphocytic Leukemia, Specified FISH,
Varies

#### Overview

#### **Useful For**

Detecting recurrent common chromosome abnormalities in patients with chronic lymphocytic leukemia (CLL)

Distinguishing patients with 11;14 translocations who have the leukemic phase of mantle cell lymphoma from patients who have CLL

Detecting patients with atypical CLL with translocations between IGH and BCL3

Evaluating specimens in which chromosome studies are unsuccessful

#### **Reflex Tests**

Test Id	Reporting Name	Available Separately	Always Performed
CLLMB	Probe, Each Additional	No, (Bill Only)	No
	(CLLMF)		

#### **Testing Algorithm**

This test includes a charge for the probe application, analysis, and professional interpretation of results for one probe set (2 individual fluorescence in situ hybridization [FISH] probes). Additional charges will be incurred for additional probe sets performed.

This is not intended as a panel test and the desired probes must be specified upon order. If the patient is being evaluated for known abnormalities, targeted probes must be listed in the probe request field.

When specified, any of the following probes will be performed:

6q-, D6Z1/MYB

11q-, D11Z1/ATM

+12, D12Z3/MDM2

13q-, D13S319/LAMP1

17p-, TP53/D17Z1

t(11;14), CCND1/IGH

t(14;19), IGH/BCL3

Appropriate ancillary probes may be performed at consultant discretion to render comprehensive assessment. Any additional probes will have the results included within the final report and will be performed at an additional charge.

#### **Method Name**

Fluorescence In Situ Hybridization (FISH)

## **NY State Available**



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Yes

## Specimen

## **Specimen Type**

Varies

## **Ordering Guidance**

This test is intended for instances when limited chronic lymphocytic leukemia (CLL) fluorescence in situ hybridization (FISH) probes are needed based on specific abnormalities or abnormalities identified in the diagnostic sample.

If targeted FISH probes are not included with this test order, test processing will be delayed and the test may be canceled and automatically reordered by the laboratory as CLLDF / Chronic Lymphocytic Leukemia, Diagnostic FISH, Varies.

If a paraffin-embedded tissue sample is received, this test will be canceled and automatically reordered by the laboratory as SLL / Small Lymphocytic Lymphoma, FISH, Tissue.

## **Shipping Instructions**

Advise Express Mail or equivalent if not on courier service.

## **Necessary Information**

A reason for testing and a flow cytometry and/or a bone marrow pathology report should be submitted with each specimen. The laboratory will not reject testing if this information is not provided; however, appropriate testing and/or interpretation may be compromised or delayed in some instances. If not provided, an appropriate indication for testing may be entered by Mayo Clinic Laboratories.

### **Specimen Required**

Submit only 1 of the following specimens:

**Preferred:** 

Specimen Type: Blood Container/Tube:

Preferred: Yellow top (ACD)

Acceptable: Green top (heparin) or lavender top (EDTA)

**Specimen Volume:** 6 mL Collection Instructions:

1. Invert several times to mix blood.

2. Send whole blood in original tube. Do not aliquot.

Acceptable:

Specimen Type: Bone marrow



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**Container/Tube:** 

Preferred: Yellow top (ACD)

Acceptable: Green top (heparin) or lavender top (EDTA)

**Specimen Volume:** 2 to 3 mL **Collection Instructions:** 

1. It is preferable to send the first aspirate from the bone marrow collection.

2. Invert several times to mix bone marrow.

3. Send bone marrow in original tube. Do not aliquot.

#### Forms

If not ordering electronically, complete, print, and send a <u>Hematopathology/Cytogenetics Test Request</u> (T726) with the specimen.

#### **Specimen Minimum Volume**

Blood: 2 mL; Bone marrow: 1 mL

#### Reject Due To

All specimens will be evaluated at Mayo Clinic Laboratories for test suitability.

## **Specimen Stability Information**

Specimen Type	Temperature	Time	Special Container
Varies	Ambient (preferred)		
	Refrigerated		

## Clinical & Interpretive

#### **Clinical Information**

Chronic lymphocytic leukemia (CLL) is one of the most common leukemias in adults. The most frequently seen cytogenetic abnormalities in CLL involve chromosomes 6, 11, 12, 13, and 17. These are detected and quantified using the CLL <u>fluorescence in situ hybridization</u> (FISH) panel.

Use of CpG-oligonucleotide mitogen will identify an abnormal CLL karyotype in at least 80% of cases. This mitogen is added to cultures when chromosome analysis is ordered and the reason for testing is B-cell lymphoproliferative disorders (CHRBM / Chromosome Analysis, Hematologic Disorders, Bone Marrow and CHRHB / Chromosome Analysis, Hematologic Disorders, Blood).

This FISH test detects an abnormal clone in approximately 70% of patients with indolent disease and greater than 80% of patients who require treatment. At least 5% of patients referred for CLL FISH testing have translocations involving the *IGH* locus. Fusion of *IGH* with *CCND1* is associated with t(11;14)(q13;q32), and fusion of *IGH* with *BCL3* is associated with t(14;19)(q32;q13.3). Patients with t(11;14) usually have the leukemic phase of mantle cell lymphoma. Patients with t(14;19) may have an atypical form of B-CLL or the leukemic phase of a lymphoma. The prognostic associations for chromosome abnormalities detected by this FISH assay are, from best to worst: 13q-, normal, +12, 6q-, 11q-, and 17p-.



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#### Reference Values

An interpretive report will be provided.

#### Interpretation

A neoplastic clone is detected when the percent of cells with an abnormality exceeds the normal reference range for any given probe set.

The absence of an abnormal clone does not rule out the presence of a chronic lymphocytic leukemia clone or another neoplastic disorder.

#### **Cautions**

This test is not approved by the US Food and Drug Administration, and it is best used as an adjunct to existing clinical and pathologic information.

#### Supportive Data

Each probe was independently tested and verified on unstimulated peripheral blood and bone marrow specimens. Normal cutoffs were calculated based on the results of at least 25 normal specimens. In addition, each probe set was evaluated in a blinded fashion to confirm the probe set detected the abnormality it was designed to detect.

#### Clinical Reference

- 1. Dewald GW, Brockman SR, Paternoster SF, et al. Chromosome anomalies detected by interphase FISH: correlation with significant biological features of B-cell chronic lymphocytic leukemia. Br J Haematol. 2003;121:287-295
- 2. Dohner H, Stilgenbauer S, Benner A, et al. Genomic aberrations and survival in chronic lymphocytic leukemia. N Engl J Med. 2000;343(26):1910-1916
- 3. Van Dyke DL, Shanafelt TD, Call TG, et al. A comprehensive evaluation of the prognostic significance of 13q deletions in patients with B-chronic lymphocytic leukaemia. Br J Haematol. 2010;148:544-550
- 4. Shanafelt TD. Predicting clinical outcome in CLL: how and why. Hematology Am Soc Hematol Educ Program. 2009;421-429
- 5. Van Dyke DL, Werner L, Rassenti LZ, et al. The Dohner fluorescence in situ hybridization prognostic classification of chronic lymphocytic leukaemia (CLL): the CLL Research Consortium experience. Br J Haematol. 2016;173(1):105-113
- 6. Fang H, Reichard KK, Rabe KG, et al. IGH translocations in chronic lymphocytic leukemia: Clinicopathologic features and clinical outcomes. Am J Hematol. 2019;94(3):338-345
- 7. Huh YO, Schweighofer CD, Ketterling RP, et al. Chronic lymphocytic leukemia with t(14;19)(q32;q13) is characterized by atypical morphologic and immunophenotypic features and distinctive genetic features. Am J Clin Pathol. 2011;135(5):686-696

## Performance

## **Method Description**

This test is performed using commercially available and laboratory-developed probes. Deletion of chromosomes 6q, 11q, 13q, and 17p, and trisomy of chromosome 12 are detected using enumeration strategy probes. A dual-color, dual-fusion fluorescence in situ hybridization (D-FISH) strategy probe set is used to detect *CCND1::IGH* rearrangements and for reflex testing to identify *IGH*::*BCL3* rearrangements. For enumeration strategy probe sets, 100 interphase nuclei are



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scored; 200 interphase nuclei are scored when D-FISH probes are used. All results are expressed as the percent abnormal nuclei.(Unpublished Mayo method)

## **PDF Report**

No

## Day(s) Performed

Monday through Friday

### **Report Available**

7 to 10 days

## **Specimen Retention Time**

4 weeks

## **Performing Laboratory Location**

Mayo Clinic Laboratories - Rochester Main Campus

#### Fees & Codes

#### Fees

- Authorized users can sign in to <u>Test Prices</u> for detailed fee information.
- Clients without access to Test Prices can contact <u>Customer Service</u> 24 hours a day, seven days a week.
- Prospective clients should contact their account representative. For assistance, contact <u>Customer Service</u>.

## **Test Classification**

This test was developed and its performance characteristics determined by Mayo Clinic in a manner consistent with CLIA requirements. It has not been cleared or approved by the US Food and Drug Administration.

#### **CPT Code Information**

88271x2, 88275, 88291-FISH Probe, Analysis, Interpretation; 1 probe set 88271x2, 88275-FISH Probe, Analysis; each additional probe set (if appropriate)

## **LOINC®** Information

Test ID	Test Order Name	Order LOINC® Value
CLLMF	CLL, Specified FISH	101920-7

Result ID	Test Result Name	Result LOINC® Value
610725	Result Summary	50397-9
610726	Interpretation	69965-2
610727	Result Table	93356-4
610728	Result	62356-1



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GC091	Reason for Referral	42349-1
GC092	Specimen	31208-2
610729	Source	31208-2
610730	Method	85069-3
610731	Additional Information	48767-8
610732	Disclaimer	62364-5
610733	Released by	18771-6
GC093	Probes Requested	78040-3