

## Overview

### Useful For

Interpretation to aid in the diagnosis of tularemia caused by *Francisella tularensis*

### Method Name

Only orderable as part of a profile. For more information see TULAB / *Francisella tularensis* Antibody, IgM and IgG, ELISA, Serum.

Technical Interpretation

### NY State Available

Yes

## Specimen

### Specimen Type

Serum

### Specimen Required

Only orderable as part of a profile. For more information see TULAB / *Francisella tularensis* Antibody, IgM and IgG, ELISA, Serum.

### Reject Due To

### Specimen Stability Information

| Specimen Type | Temperature              | Time    | Special Container |
|---------------|--------------------------|---------|-------------------|
| Serum         | Refrigerated (preferred) | 7 days  |                   |
|               | Frozen                   | 30 days |                   |

## Clinical & Interpretive

### Clinical Information

*Francisella tularensis* is a small, intracellular, coccobacillary gram-negative bacterium and is an obligate pathogen in animals and humans, primarily maintained in rabbits, hares, cats, ticks, and deerflies. *F tularensis* is found throughout North America and parts of Asia and, similar to *Brucella* species, is considered a potential agent of bioterrorism. Human infection with *F tularensis* usually occurs through inhalation of infected aerosols, ingestion of contaminated meat or

water, handling of diseased or sick animals, or through the bite of an infected arthropod (eg, tick, deerflies).

Following a 3- to 5-day incubation period, the clinical manifestations of infection with *F tularensis* differ primarily depending on the site and route of infection. The most common form of disease is ulceroglandular (45%-80% of cases), which is associated with an arthropod (or animal) bite or another cause of skin barrier compromise. This leads to development of a painful papule that ultimately ulcerates allowing the bacterium to enter the lymphatic system. Glandular tularemia is similar in presentation to ulceroglandular disease; however, it lacks the ulceration and, more frequently, causes septicemia. Other, less frequent clinical manifestations include oculoglandular (Parinaud syndrome), oropharyngeal and gastrointestinal disease, and pneumonic or typhoidal tularemia.

Diagnostic testing options for *F tularensis* primarily include culture and serology. Providers suspecting tularemia should collect appropriate specimens (eg, skin lesion biopsy, lymph node aspirates) promptly and send for culture. The microbiology laboratory should be alerted to the possibility of *F tularensis* to ensure that appropriate safety measures are taken to protect the laboratory technologists. Growth on culture is a definitive means of making a diagnosis of tularemia. Serologic testing may be used to support a diagnosis of current or recent tularemia in patients who are IgM positive, who seroconvert to IgM, or who are IgG positive in paired sera collected 2 to 3 weeks apart.

## Reference Values

Only orderable as part of a profile. For more information see TULAB / *Francisella tularensis* Antibody, IgM and IgG, ELISA, Serum.

IgM: Negative

IgG: Negative

Reference values apply to all ages.

## Interpretation

| IgM result | IgG result | Interpretation   |
|------------|------------|--|
| Negative   | Negative   | No antibodies to <i>Francisella tularensis</i> detected. Antibody response may be negative in samples collected too soon following infection/exposure. Repeat testing on a new sample in 1 to 2 weeks if clinically indicated. |
| Positive   | Negative   | IgM class antibodies to <i>F tularensis</i> detected, suggesting current or recent infection. Repeat testing in 1 to 2 weeks to detect seroconversion of IgG may be considered to confirm the diagnosis.                       |
| Positive   | Borderline |  |
| Borderline | Negative   | Questionable presence of IgM antibodies to <i>F tularensis</i> . Consider repeat testing in 1 to 2 weeks.  |
| Borderline | Positive   | IgG class antibodies to <i>F tularensis</i> detected suggesting recent or past infection. Clinical correlation alongside presentation, exposure history and other laboratory findings required.                                |

|            |            |  |
|------------|------------|--|
| Borderline | Borderline | Questionable presence of IgM and IgG class antibodies to <i>F tularensis</i> . Consider repeat testing in 1 to 2 weeks.  |
| Positive   | Positive   | IgM and IgG class antibodies to <i>F tularensis</i> detected suggesting current, recent or past infection. Cross-reactions may occur in patients with a current or prior Brucella infection. Clinical correlation alongside presentation, exposure history and other laboratory findings required. |
| Negative   | Positive   | IgG class antibodies to <i>F tularensis</i> detected suggesting recent or past infection. Clinical correlation alongside presentation, exposure history and other laboratory findings required.  |
| Negative   | Borderline | Questionable presence of IgG antibodies to <i>F tularensis</i> . Consider repeat testing in 1 to 2 weeks.  |

### Cautions

False-negative results may occur in specimens collected too soon following symptom onset, prior to the development of a detectable immune response. Repeat testing on new specimens collected 2 to 4 weeks later may be helpful.

False-positive results may occur in patients previously or currently infected with *Brucella* species. Other less frequent causes of cross-reactivity that have been reported include prior infection with *Yersinia*, *Salmonella*, or *Legionella* species.

IgM-class antibodies may be detectable as soon as 1 week after symptom onset and may remain detectable for multiple years following resolution of disease in some individuals. Therefore, an IgM positive result may not indicate current or recent infection in some cases.

Multiple subspecies of *Francisella tularensis*, including *F tularensis* subspecies *tularensis*, *F tularensis* subspecies *holarctica*, and *F tularensis* subspecies *novicida* are found throughout the northern hemisphere, including in the United States. The IgM and IgG anti-*F tularensis* [enzyme-linked immunosorbent assays](#) used at Mayo Clinic Laboratories are based on the lipopolysaccharide (LPS) antigen of *F tularensis*. Although not directly tested, previous studies indicate that there are no antigenic differences between the LPS of *F tularensis* subspecies *tularensis* and the other subspecies. Therefore, these assays should not be used to differentiate between infection with the various *F tularensis* subspecies.

### Clinical Reference

- Petersen JM, Schriefer ME, Araj GE. Francisella and Brucella. In: Carroll KC, Pfaller MA, Landry ML, et al, eds. Manual of Clinical Microbiology. 12th ed. AMS Press; 2019
- Nigrovic LE, Wingarter SL. Tularemia. Infect Dis Clin North Am. 2008;22(3):489-504. doi:10.1016/j.idc.2008.03.004

### Performance

### Method Description

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Automated interpretation of IgM and IgG antibody results for *Francisella tularensis*.

**PDF Report**

No

**Day(s) Performed**

Tuesday, Thursday

**Report Available**

Same day/1 day

**Performing Laboratory Location**

Mayo Clinic Laboratories - Rochester Superior Drive

## Fees & Codes

**Fees**

- Authorized users can sign in to [Test Prices](#) for detailed fee information.
- Clients without access to Test Prices can contact [Customer Service](#) 24 hours a day, seven days a week.
- Prospective clients should contact their account representative. For assistance, contact [Customer Service](#).

**Test Classification**

Not Applicable

**LOINC® Information**

| Test ID | Test Order Name              | Order LOINC® Value |
|---------|------------------------------|--------------------|
| TULI    | F. tularensis Interpretation | 93718-5            |

| Result ID | Test Result Name             | Result LOINC® Value |
|-----------|------------------------------|---------------------|
| TULI      | F. tularensis Interpretation | 93718-5             |