

# Inflammatory Myofibroblastic Tumors (IMT), 2p23 (ALK) Rearrangement, FISH, Tissue

# Overview

#### Useful For

Supporting the diagnosis of inflammatory myofibroblastic tumors when used conjunction with an anatomic pathology consultation

#### **Reflex Tests**

Test Id	Reporting Name	Available Separately	Always Performed
_PBCT	Probe, +2	No, (Bill Only)	No
_PADD	Probe, +1	No, (Bill Only)	No
_PB02	Probe, +2	No, (Bill Only)	No
_PB03	Probe, +3	No, (Bill Only)	No
_IL25	Interphases, <25	No, (Bill Only)	No
_1099	Interphases, 25-99	No, (Bill Only)	No
_1300	Interphases, >=100	No, (Bill Only)	No

#### **Testing Algorithm**

This test includes a charge for the probe application, analysis, and professional interpretation of results for one probe set (2 individual fluorescence in situ hybridization probes). No analysis charges will be incurred if an insufficient number of representative cells are available for analysis.

Appropriate ancillary probes may be performed at consultant discretion to render comprehensive assessment. Any additional probes will have the results included within the final report and will be performed at an additional charge.

#### Method Name

Fluorescence In Situ Hybridization (FISH)

#### **NY State Available**

Yes

## Specimen

Specimen Type

Tissue

#### **Ordering Guidance**

This test does not include a pathology consultation. If a pathology consultation is requested, order PATHC / Pathology Consultation, and appropriate testing will be added at the discretion of the pathologist and performed at an additional



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charge.

Multiple oncology (cancer) gene panels are also available. For more information see <u>Hematology, Oncology, and</u> <u>Hereditary Test Selection Guide</u>.

# Additional Testing Requirements

Confirmation testing by next-generation sequencing to resolve atypical or unbalanced fluorescence in situ hybridization results of this gene region is available, order SARCP / Sarcoma Targeted Gene Fusion/Rearrangement Panel, Next-Generation Sequencing, Tumor.

## **Shipping Instructions**

Advise Express Mail or equivalent if not on courier service.

# Necessary Information

**1.** A pathology report is required for testing to be performed. If not provided, appropriate testing and/or interpretation may be compromised or delayed. Acceptable pathology reports include working drafts, preliminary pathology, or surgical pathology reports.

## 2. The following information must be included in the report provided.

- 1. Patient name
- 2. Block number must be on all blocks, slides, and paperwork
- 3. Date of collection
- 4. Tissue Source

**3.** A reason for testing must be provided. If this information is not provided, an appropriate indication for testing may be entered by Mayo Clinic Laboratories.

## **Specimen Required**

Submit only 1 of the following specimens:

## Preferred

## Specimen Type: Tissue block

**Collection Instructions:** Submit a formalin-fixed, paraffin-embedded (FFPE) tumor tissue block. Blocks prepared with alternative fixation methods will be attempted but are less favorable for successful results by FISH testing; provide fixation method used.

## Additional Information:

- 1. Paraffin-embedded specimens can be from any anatomic location (skin, soft tissue, lymph node, etc).
- 2. Bone specimens that have been decalcified will be attempted for testing, but the success rate is approximately 50%.

## Acceptable

Specimen Type: Tissue slides

Slides: 1 Hematoxylin and eosin stained and 4 unstained

**Collection Instructions**: Submit 4 consecutive unstained, positively charged, unbaked slides with 5 micron-thick sections of the tumor tissue and 1 slide stained with hematoxylin and eosin.

## Forms

If not ordering electronically, complete, print, and send an <u>Oncology Test Request</u> (T729) with the specimen.



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#### **Specimen Minimum Volume**

Slides: 1 Hematoxylin and eosin stained and 2 unstained

#### **Reject Due To**

All specimens will be evaluated at Mayo Clinic Laboratories for test suitability.

#### **Specimen Stability Information**

Specimen Type	Temperature	Time	Special Container
Tissue	Ambient (preferred)		
	Refrigerated		

## Clinical & Interpretive

#### **Clinical Information**

Inflammatory myofibroblastic tumor (IMT) is a distinctive lesion composed of myofibroblastic spindle cells accompanied by an inflammatory infiltrate which occur primarily in the soft tissue but may arise in any anatomical site including lung, soft tissue, retroperitoneum, and bladder.

A subset of IMT are characterized by rearrangements involving the *ALK* gene at 2p23. Studies support that identification of *ALK* gene rearrangement is useful to differentiate IMTs from other spindle cell neoplasms of soft tissue and viscera.

## **Reference Values**

An interpretive report will be provided.

#### Interpretation

ALK will be clinically interpreted as positive or negative.

A neoplastic clone is detected when the percent of cells with an abnormality exceeds the normal cutoff for the ALK probe set.

A positive result is consistent with rearrangement of the *ALK* gene and likely reflects *ALK* fusion with a partner gene. The significance of this finding is dependent on the clinical and pathologic features.

A negative result suggests an ALK gene rearrangement is not present but does not completely exclude the presence of an ALK gene rearrangement and does not exclude the diagnosis of inflammatory myofibroblastic tumors.

#### Cautions

This test is not approved by the US Food and Drug Administration and it is best used as an adjunct to existing clinical and pathologic information.

This fluorescence in situ hybridization (FISH) assay does not rule out other chromosome abnormalities.



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Fixatives other than formalin (eg, Prefer, Bouin's) may not be successful for FISH assays. Non-formalin fixed specimens will not be rejected.

Paraffin-embedded tissues that have been decalcified may not be successful for FISH analysis. The success rate of FISH studies on decalcified tissue is approximately 50%, but FISH will be attempted if sufficient tumor is present for analysis.

FISH studies will be attempted if sufficient tumor is present for analysis. The pathologist reviewing the hematoxylin and eosin-stained slide may find it necessary to cancel testing if insufficient tissue/tumor is available for testing.

If no FISH signals are observed post-hybridization, the case will be released indicating a lack of FISH results.

## **Clinical Reference**

1. Sukov WR, Cheville JC, Carlson AW, et al. Utility of ALK-1 protein expression and ALK rearrangements in distinguishing inflammatory myofibroblastic tumor from malignant spindle cell lesions of the urinary bladder. Mod Pathol. 2007;20(5):592-603

2. Tsuzuki T, Magi-Galluzzi C, Epstein JI. ALK-1 expression in inflammatory myofibroblastic tumor of the urinary bladder. Am J Surg Pathol. 2004;28(12):1609-1614

3. WHO Classification of Tumours Editorial Board. Soft Tissue and Bone Tumours. 5th ed. IARC; 2020. WHO Classification of Tumours Series. Vol. 3, 109-111

# Performance

## **Method Description**

The test is performed using a commercially available *ALK* dual-color break-apart strategy probe (BAP). Formalin-fixed, paraffin-embedded tissues are cut at 5 microns and mounted on positively charged glass slides. The selection of tissue and the identification of target areas on the hematoxylin and eosin (H and E)-stained slide is performed by a pathologist. Using the H and E-stained slide as a reference, target areas are etched with a diamond-tipped etcher on the back of the unstained slide to be assayed. The probe set is hybridized to the appropriate target areas and 2 technologists each analyze 50 interphase nuclei (100 total) with the results expressed as the percent of abnormal nuclei.(Unpublished Mayo method)\

## **PDF Report**

No

# Day(s) Performed

Monday through Sunday

## **Report Available**

7 to 10 days

## **Specimen Retention Time**

Slides and H&E used for analysis are retained by the laboratory in accordance with regulatory requirements. Client provided paraffin blocks and extra unstained slides (if provided) will be returned after testing is complete.



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# Performing Laboratory Location

Mayo Clinic Laboratories - Rochester Main Campus

# Fees & Codes

#### Fees

- Authorized users can sign in to <u>Test Prices</u> for detailed fee information.
- Clients without access to Test Prices can contact <u>Customer Service</u> 24 hours a day, seven days a week.
- Prospective clients should contact their account representative. For assistance, contact Customer Service.

#### **Test Classification**

This test was developed and its performance characteristics determined by Mayo Clinic in a manner consistent with CLIA requirements. It has not been cleared or approved by the US Food and Drug Administration.

## **CPT Code Information**

88271x2, 88291-DNA probe, each (first probe set), Interpretation and report
88271x2-DNA probe, each; each additional probe set (if appropriate)
88271x1-DNA probe, each; coverage for sets containing 3 probes (if appropriate)
88271x2-DNA probe, each; coverage for sets containing 4 probes (if appropriate)
88271x3-DNA probe, each; coverage for sets containing 5 probes (if appropriate)
88274 w/modifier 52-Interphase in situ hybridization, <25 cells, each probe set (if appropriate)</li>
88274-Interphase in situ hybridization, 25 to 99 cells, each probe set (if appropriate)

## LOINC<sup>®</sup> Information

Test ID	Test Order Name	Order LOINC <sup>®</sup> Value
IMTF	ALK, (2p23), IMT, FISH, Ts	78205-2
Result ID	Test Result Name	Result LOINC <sup>®</sup> Value
52171	Result Summary	50397-9
52173	Interpretation	69965-2
54587	Result	62356-1
CG747	Reason for Referral	42349-1
52174	Specimen	31208-2
52175	Source	31208-2
52176	Tissue ID	80398-1
52177	Method	85069-3
55029	Additional Information	48767-8
52178	Released By	18771-6
53828	Disclaimer	62364-5